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# Virulence of four entomopathogenic nematode against different stages of the Mediterranean fruit fly, *Ceratitis capitata* Wiedemann (Diptera: Tephritidae)

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## Abstract

**Background:** *Ceratitis capitata* Wiedemann (Diptera: Tephritidae) (Mediterranean fruit fly), is one of the important polyphagous pest species in the world that harms almost all fruits causing significant yields losses. Control of this pest which is on the quarantine list of many countries is highly important. Due to the negative effects of chemical control on the environment and human health, biological control approaches have gained importance. Entomopathogenic nematodes (EPNs) are a potential biological control agent that can be used for effectively controlling the Medfly.

**Results:** In this study, effects of four EPNs species on larvae, pupae and adults of Medfly were investigated under laboratory conditions. Four different concentrations of EPNs species were used against each stage of the pest. It pest showed different sensitivity to different concentrations of each of the tested EPNs. High mortality rate of 94% was caused by *Heterorhabditis bacteriophora* (11 KG), followed by *H. bacteriophora* (TOK-20) with 91%, *Steinernema carpocapsae* (85%) and *S. feltiae* (Tokat-Emir) with 71% at highest concentration (200 IJ/larvae). The highest effect on adult mortality (100%) was recorded by the species *S. feltiae* (Tokat-Emir) at the highest concentration (200 IJ/adult) in the trials, followed by *H. bacteriophora* (11 KG) with 92%, *H. bacteriophora* (TOK-20) with 91% and *S. carpocapsae* (Tokat-Bakışlı05) with 87.37% mortality rates at the same concentrations.

**Conclusions:** It was determined that the EPNs tested in this study had insecticidal properties and they might be used in biological control programs against the Medfly.

**Keywords:** *Ceratits capitata*, Biological control, Entomopathogenic nematodes, Efficiency

## Background

The Mediterranean fruit fly, *Ceratitis capitata* Wiedemann (Diptera: Tephritidae) is one of the economic polyphagous pest species in the world (Zucchi 2001). It has been recorded in 350 hosts from 65 different families around the world, especially to temperate climate fruits and subtropical fruits, some vegetables and ornamental plants worldwide (Weems 1981; Liquido et al. 1990, 1991;

Woods et al. 2005; Thomas et al. 2010). Its damages to the exported products is very important for the country's economy as this causes economic losses by affecting the yield quantity and quality (Başpınar et al. 2009). Recently, researchers are seeking for alternatives pest control tools like biological control agents. Entomopathogenic nematodes (EPNs) are obligate insect pathogens found in soil that are used as biological agents. They have a wide host range, able to kill their hosts with the help of their symbiotic bacteria within 24–48 h, easy to mass produce, able to actively search and find their hosts, can stay alive for a long time in the absence of hosts, and do not harm the environment and humans. Studies of isolating EPNs and

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investigating their virulences have increased, and are of great importance for biological control studies (Hazır et al. 2003). Although the potentials of EPNs against many pest groups in the world have been demonstrated in laboratory and field / garden studies, there are few studies on fruit pests.

This study aimed to determine the virulences of 4 EPNs species [*Steinernema carpocapsae* (Weiser), *S. feltiae* (Filipjev) (Rhabditida: Steinernematidae) and *Heterorhabditis bacteriophora* Poinar (Rhabditida: Heterorhabditidae)] on the larvae, pupae and adults of the Medfly under laboratory conditions.

## Methods

### Nematodes' source

The EPNs species (*Steinernema feltiae* (Tokat-Emir), *S. carpocapsae* (Tokat-Bakışlı05), *Heterorhabditis bacteriophora* (TOK-20), *H. bacteriophora* (11-KG) were obtained from the Plant Protection Department of Tokat Gaziosmanpaşa University, Turkey. Infective juveniles of the 4 species were reared on last instar larvae of the greater wax moth, *Galleria mellonella* (L.) described by Kaya and Stock (1997).

### Mass rearing of *Galleria mellonella* larvae

*Galleria mellonella*'s larvae were reared on a artificial diet, contained 890 g flour, 222 g dry baker's yeast, 500 g glycerin, 500 g honey, 445 g milk powder and 125 g bee-wax in an incubator with 16/8 h lighting set at 24–25 °C (Mohamed and Coppel 1983).

### Mass rearing of entomopathogenic nematode species

Ten last instar *G. mellonella* larvae were placed in small Petri dishes with lined Whatman paper. A suspension EPNs of infective juveniles inoculated the larvae. Petri dishes were placed in an incubator at 20–23 °C. Dead larvae was controlled frequently. Infective EPNs juveniles were obtained from infected *G. mellonella* larvae, using the "White trap" method (White 1927). Juveniles were kept in a refrigerator at +10 °C.

### Rearing of the Medfly

The stock culture of the Medfly, reared in the Entomology Laboratory, was used in the experiments. Fruit fly production was carried out in plexiglass adult cages with 50 mesh tulle on both sides in climate rooms with 25 ± 1 °C, 65 ± 5% relative humidity, and 16 h light and 8 h dark conditions. Eggs laid by mature individuals were left on large Petri dishes on artificial media made from a mixture of bran, wheat germ, yeast, hydrochloric acid and water. The last instar larvae were collected 6–8 days after the eggs laying to be used in the experiments.

## Experiments

Experiments were conducted using 150 ml plastic containers, containing the last instar larvae and pupae of the pest on soil mixture (80% sand, 15% soil and 5% clay) sterilized at 121 °C (Chen et al. 1995). Five-to 7-day-old pupae were used in the experiments. In adult trials, EPNs were applied by placing Whatman paper on the bottom of 150 ml plastic containers and a piece of cotton soaked in sugar melted water was also left in each container. Four concentrations (0, 50, 100, 200 IJs/*C. capitata* stages) for each nematode species were tested against both late instar larvae, pupae and adults of *C. capitata*. Trials had 10 replications of each concentration and 5 individuals per repeat (50 individuals in total). After placing the 5 individuals of the pest in each plastic container, the nematodes were applied by a pipette. Only distilled water and sugar melted water were provide as food for the adults in control group. All the trials were repeated twice, on different dates, and the set up was incubated at 25 °C under 16 h dark and 8 h light climatic room conditions. Mortality rates of adults and larvae were calculated 5 days after the inoculation, while pupae were calculated 10 days later. Dead medfly larvae, pupae and adults were collected and placed in the White trap method. About one week later, EPNs emerged from insect cadavers and were examined under a stereomicroscope.

### Statistical analyses

In experiments, all data were analyzed by statistical program JMP 7. Analyses were done using one-way ANOVA and used "LSMeans Student's t test" for finding the variety in the effects of different nematode species and concentrations on Medfly.

## Results

Data were evaluated 5 days after EPNs inoculation of the medfly's larvae and adults. Pupal data were evaluated after 10 days. Statistically, both parameters (species and concentrations) were found to be significant. Mortality rates of the last instar larvae of the medfly increased directly as the concentration increased in case of all EPNs species ( $p < 0.001$ ).

### Virulence of EPN species at different concentrations against *Ceratitis capitata* last instar larvae

It was observed that all the applications against the Medfly last instar larvae were quite effective than the controls. High mortality rates of all EPNs species against the last instar larvae of the Medfly were recorded at 200 IJs/larva (at high concentration). High mortality rate (94%) was reported in case of *H. bacteriophora* (11 KG) isolate, followed by *H. bacteriophora* (TOK-20) with (91%), *S.*

*carpocapsae* (Tokat-Bakışlı05) with (85%), and *S. feltiae* (Tokat-Emir) with (71%) at 200 IJs/larva. Mortality rates at the 25 IJs/larva concentration were found close to those of 200 IJs/larva. At this concentration, the highest deaths were recorded by *H. bacteriophora* (11 KG) isolate with (88%), followed by *H. bacteriophora* (TOK-20) with (87%), *S. carpocapsae* (Tokat-Bakışlı05) with (85.88%), and *S. feltiae* (Tokat-Emir) with (71%). At 100 IJs/larva concentration, *H. bacteriophora* (11 KG) was found to cause the highest mortality rate with (88%). In addition, the mortality rates in other species were determined as (87%) for *H. bacteriophora* (TOK-20), (85.88%) for *S. carpocapsae* (Tokat-Bakışlı05), and (71%) for *S. feltiae* (Tokat-Emir) at concentration of 100 IJs/larva. The lowest mortality rate in the last instar larvae of the pest was by 50 IJs/*C. capitata* larva. The mortality rates at this concentration 50 IJs/larva were in *H. bacteriophora* (11 KG) isolate with (87%), followed by *H. bacteriophora* (TOK-20) with (84%), *S. feltiae* (Tokat-Emir) with (66%) and *S. carpocapsae* (Tokat-Bakışlı05) with (64%) (Table 1).

#### Virulence of EPN species at different concentrations against pupae of *C. capitata*

The pupae were evaluated at 10 days after from the application. While the difference between the concentrations was significant, the difference between the species was not statistically significant. In pupal stage trials, applications were found to be effective than the control. The highest mortality rate was found at 200 IJs/pupa concentrations for all EPNs nematode species.

*S. carpocapsae* (Tokat-Bakışlı05) isolate with (52%), followed by *H. bacteriophora* (11 KG) with (51%), *H. bacteriophora* (TOK-20) with (49%), *S. feltiae* (Tokat-Emir) with (43%) at 200 IJs/pupa (Table 2). The least mortality rate was calculated 50 IJs/pupa concentrations to all nematode species.

#### Virulence of EPN species at different concentrations against adults of *Ceratitis capitata*

In all trials and concentrations against adults of the pest were found to be more effective than controls. In addition, as the concentration increased in all EPNs species used and the mortality rate of the adult individuals increased in direct proportion ( $p < 0.001$ ).

The highest mortality rate (100%) detected at 200 IJs/adult concentration against the adults, was caused by *S. feltiae* (Tokat-Emir), followed by *H. bacteriophora* (11 KG) with (92%), *H. bacteriophora* (TOK-20) with (91%), and *S. carpocapsae* (Perspective 05) with (87.37%). At 100 IJs/adult concentration, the highest deaths were observed in *S. feltiae* (Tokat-Emir) with (89.47%), followed by *H. bacteriophora* (11 KG) with (88%), *H. bacteriophora* (TOK-20) with (86%), and (61.05%) for *S. carpocapsae* (Bakışlı 05). The lowest mortality rate in adults of *C. capitata* was at 50 IJs/adult concentration. The mortality rates at 50 IJs/adult were (84%) with *H. bacteriophora* (TOK-20), (83%) with *S. feltiae* (Tokat-Emir), (78%) with *S. carpocapsae* (Bakışlı 05) and (75.56%) with *H. bacteriophora* (11 KG) isolates (Table 3).

**Table 1** Effect of nematode species and concentrations on mortality rate of Mediterranean fruit fly larvae

| Nematode species                                 | Concentrations (IJs/larvae) |                          |                           |                           |
|--|-----------------------------|--------------------------|---------------------------|---------------------------|
|  | Control                     | 50IJ/ <i>C. capitata</i> | 100IJ/ <i>C. capitata</i> | 200IJ/ <i>C. capitata</i> |
| <i>Heterorhabditis bacteriophora</i> (11 KG)     | 12.00 <sup>d</sup>          | 87.00 <sup>ab</sup>      | 88.00 <sup>ab</sup>       | 94.00 <sup>a</sup>        |
| <i>H. bacteriophora</i> (TOK-20)                 | 9.00 <sup>d</sup>           | 84.00 <sup>b</sup>       | 87.00 <sup>ab</sup>       | 91.00 <sup>ab</sup>       |
| <i>Steinernema carpocapsae</i> (Tokat-Bakışlı05) | 12.00 <sup>d</sup>          | 64.00 <sup>c</sup>       | 85.88 <sup>ab</sup>       | 85.00 <sup>ab</sup>       |
| <i>S. feltiae</i> (Tokat-Emir)                   | 10.00 <sup>d</sup>          | 66.00 <sup>c</sup>       | 71.00 <sup>c</sup>        | 71.00 <sup>c</sup>        |

CV: 0.23, LSD: 9.10

Means followed by the same letter are not statistically different according to the Duncan test ( $P \leq 0.05$ )

**Table 2** Effect of nematode species and concentrations on mortality rates of Medfly pupae

| Nematode species                                 | Concentrations (IJs/pupa) |                          |                           |                           |
|--|---------------------------|--------------------------|---------------------------|---------------------------|
|  | Control                   | 50IJ/ <i>C. capitata</i> | 100IJ/ <i>C. capitata</i> | 200IJ/ <i>C. capitata</i> |
| <i>Heterorhabditis bacteriophora</i> (11 KG)     | 10.00 <sup>d</sup>        | 32.00 <sup>c</sup>       | 42.00 <sup>b</sup>        | 51.00 <sup>a</sup>        |
| <i>H. bacteriophora</i> (TOK-20)                 | 6.00 <sup>d</sup>         | 26.00 <sup>c</sup>       | 40.00 <sup>b</sup>        | 49.00 <sup>a</sup>        |
| <i>Steinernema carpocapsae</i> (Tokat-Bakışlı05) | 5.00 <sup>d</sup>         | 32.00 <sup>c</sup>       | 42.00 <sup>b</sup>        | 52.00 <sup>a</sup>        |
| <i>S. feltiae</i> (Tokat-Emir)                   | 12.00 <sup>d</sup>        | 25.00 <sup>c</sup>       | 32.00 <sup>b</sup>        | 43.00 <sup>a</sup>        |

CV: 0.56, LSD: 5.47

Means followed by the same letter are not statistically different according to the Duncan test ( $P \leq 0.05$ )

**Table 3** Effect of nematode species and concentrations on mortality rate of Medfly's adults

| Nematode species                                 | Concentrations (IJs/adult) |                          |                           |                           |
|--|----------------------------|--------------------------|---------------------------|---------------------------|
|  | Control                    | 50IJ/ <i>C. capitata</i> | 100IJ/ <i>C. capitata</i> | 200IJ/ <i>C. capitata</i> |
| <i>Heterorhabditis bacteriophora</i> (11 KG)     | 6.00 <sup>f</sup>          | 75.56 <sup>d</sup>       | 89.00 <sup>b</sup>        | 92.00 <sup>ab</sup>       |
| <i>H. bacteriophora</i> (TOK-20)                 | 7.00 <sup>f</sup>          | 84.00 <sup>bd</sup>      | 86.00 <sup>bc</sup>       | 91.00 <sup>ab</sup>       |
| <i>Steinernema carpocapsae</i> (Tokat-Bakışlı05) | 10.00 <sup>f</sup>         | 78.89 <sup>cd</sup>      | 61.05 <sup>e</sup>        | 87.37 <sup>bc</sup>       |
| <i>S. feltiae</i> (Tokat-Emir)                   | 9.00 <sup>f</sup>          | 83.00 <sup>bd</sup>      | 89.47 <sup>b</sup>        | 100 <sup>a</sup>          |

CV 0.23, LSD: 9.42

Means followed by the same letter are not statistically different according to the Duncan test ( $P \leq 0.05$ )

## Discussion

*C. capitata* is a highly invasive species. It has a high dispersive ability, a very large host range. It has a high economic impact, affecting production, control costs and market access. (Malacrida et al. 2007). Several studies have been carried out to evaluate the efficiency of steinernematid and heterorhabditid nematodes against *C. capitata*. Karagöz et al. (2009) showed that *Steinernema weiseri*, *S. feltiae*, *S. carpocapsae* and 2 strains of *Heterorhabditis bacteriophora*, isolated from Turkish soils, were very effective against larvae of the Medfly. On the other hand, Minas et al. (2016) tested 8 strains, 6 of them were native to Brazil, in sand columns against 3rd instar larvae (L3) of *C. capitata*. The highest mean mortality values reached by *Heterorhabditis* sp. LPP17, *Heterorhabditis* sp. LPP14 and *H. baujardi* LPP7 were 98.5, 95.5 and 90%, respectively. In another study Malan and Manrakhan (2009) *H. bacteriophora*, *H. zealandica* and *S. khoisanae* to infect pupariating larvae, pupae and adults of *C. capitata* and *Ceratitis rosa* was investigated in laboratory bioassays. Significantly more larvae of *C. capitata* were infected by *H. bacteriophora*. For *C. rosa*, the highest infectivity of larvae was obtained by *H. zealandica*. In contrast, adults of both species were highly infected by *S. khoisanae*. Pupariating larvae and adult flies were susceptible to all nematode infection, with no infection was recorded for the pupae. Rohde et al. (2020) reported that *Steinernema carpocapsae* ALL and *Heterorhabditis amazonensis* JPM4 were found effective in controlling larvae and pupae of *C. capitata*. Both nematodes killed *C. capitata* larvae and pupae, but *S. carpocapsae* ALL was the most effective. The all evaluated concentrations of *H. amazonensis* JPM4 nematodes also killed *C. capitata* larvae and pupae at different ages. *H. amazonensis* JPM4 nematodes effective with 28 to 54% for young pupae and 11 to 44% for old pupae. In Morocco, Mokrini et al. (2020) evaluated the efficiency of EPN strains against *C. capitata*. In laboratory assays, *S. feltiae*-SF-MOR9, *S. feltiae*-SF-MOR10 and *H. bacteriophora*-HB-MOR7 strains showed significantly higher infectivity

and penetration rates than other strains. *S. feltiae*-SF-MOR9 caused the highest larval mortality rate (80%) at 50 infective juveniles (IJs). Gazit et al. (2010) tested 12 EPN strains. *Steinernema riobrave* Texas (Sr TX) and *Heterorhabditis* sp. IS-5 (H IS-5) showed high activity and induced >80% mortality. Six EPN strains showed limited activity (>30% mortality), and 4 strains had no effect (<20% mortality). Soliman et al. (2014) reported that the ability of *H. bacteriophora* infective juveniles to infect last instar larvae of *B. zonata* and *C. capitata* was more than *S. riobrave*.

## Conclusions

Four EPNs species were found very effective against the *C. capitata* last instar larvae and adults. *S. feltiae* (Tokat-Emir) provided to be the best candidate among the other EPN species to control Medfly larvae, while *H. bacteriophora* (11 KG) was the best among the other EPN species to control the adults. However, the 4 isolates could all be used for the control of *C. capitata* biological control program in orchards. However, more field studies are needed to be conducted.

## Abbreviations

EPNs: Entomopathogenic nematodes; IJs: Infective juveniles; *C. capitata* *Ceratitis capitata*; *Cydia pomonella*; *H. bacteriophora*: *Heterorhabditis bacteriophora*; *S. carpocapsae*: *Steinernema carpocapsae*; *S. feltiae*: *Steinernema feltiae*; *G. mellonella*: *Galleria mellonella*; Medfly: Mediterranean fruit fly.

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## Authors' contributions

M.Y participated in setting the work planning and executing the experimental work. T.A.F rearing *C. capitata* larvae and adults participated in experimental studies. F.D.E and M.Y mass rearing entomopathogenic nematodes and participated in experimental studies. M.Ş analyzed the all data (statistical analyses) in study. All authors read and approved the final manuscript.

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#### Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

#### Declarations

#### Ethics approval and consent to participate

Not applicable.

#### Consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no competing interest.

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