RESEARCH Open Access

Controlling the western flower thrips, Frankliniella occidentalis (Pergande) (Thysanoptera: Thripidae) by releasing the predatory phytoseiid mites and pesticides on pepper in a greenhouse



Ahmad S. Sanad¹ and Gamal M. Hassan^{2*}

Abstract

Two programs for controlling the western flower thrips (WFT), *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae), were evaluated. Greenhouse experiments were conducted for reducing *F. occidentalis* populations on pepper plants. The first program was by releasing four phytoseiid predacious mite species namely *Neoseiulus arundonaxi* (Metwally and Sanad), *N. barkeri* (Hughes), *N. bicaudus* (Wainstein), and *Cydnoseius negevi* (Swirski and Amitai) in the presence of blue sticky traps. The second one was by applying four sequential applications of different pesticides. Results showed that two releases of *N. bicaudus*, *N. barkeri*, and *N. arundonaxi* in the presence of the blue sticky traps reduced WFT population by 45 to 56%. The best control management of the WFT was by the following insecticides: chlorfenapyr, imidacloprid, azadirachtin, and agricultural potassium soap (third program).

Keywords: Frankliniella occidentalis, Predatory phytoseiid mites, Insecticides, Blue sticky trap, Biological control, Pepper

Background

Some abiotic and biotic factors affect pepper production under greenhouse conditions. One of the biotic effects is the western flower thrips (WFT), *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae). The pest attacks many vegetable plants (Kirişik and Erler 2017). *F. occidentalis* is characterized by its small size, highly reproductive, and spread capability causing a high infestation rate (Cloyd 2009). The pest deposits its eggs in plant tissue and pupates in the soil or plant litter surrounding plant on the ground, while nymphs and adults inhabit flowers. In a severe infestation, the appropriate pest management was conducted by multiple applications of insecticides (Bethke et al. 2010).

In Egypt, Shalaby (2015) studied the population fluctuation of *F. occidentalis* on pepper flowers in a

greenhouse, while Hanafy (2015) studied its host pre-

²Vegetable, Medicinal, Aromatic and Ornamental Plant Pests Department, Plant Protection Research Institute, Agricultural Research Center, Giza, Egypt Full list of author information is available at the end of the article



ference and control techniques on cucumber, squash, eggplant, and kidney bean crops. He indicated that acetamiprid and thiamethoxam gave a high reduction in F. occidentalis population on cucumber. Multiuse of synthetic pesticides had led to a resistance against major insecticide groups (Demirozer et al. 2012). The potential of *F. occidentalis* was developed extremely fast resistance against neonicotinoid insecticides (Minakuchi et al. 2013). The resistance had been stated for chlorpyrifos and dimethoate (Robb et al. 1995) and against permethrin, methomyl, abamectin, bifenthrin, and chlorpyrifos (Immaraju et al. 1992). The insecticide residue and toxicity problems on marketable pepper crops, towards beneficial organisms (non-target species) and environmental contamination, generated the need of alternative methods to control WFT. Apart from insecticide control, biocontrol agents such as bugs, Orius spp. (Hemiptera: Anthocoridae) as well as plant extracts, are proved

^{*} Correspondence: dr.jimy.hassan@gmail.com

to be suitable and effective methods for *F. occidentalis* control (Arthurs et al. 2009 and Dogramaci et al. 2011).

Several species of the family Phytoseiidae (Acari: Mesostigmata) are important predatory mites on many crops. Numerous phytoseiid species had been reported as effective predators of WFT such as *Neoseiulus cucumeris* (Oudemans), *Amblyseius swirskii* (Athias-Henriot), and *Amblydromalus limonicus* (Garman and McGregor) (Messelink et al. 2006 and Knapp et al. 2013). *Neoseiulus barkeri* (Hughes) was commonly used as a biological control agent for thrips in pepper (Ramakers 1988). Moreover, few studies had conducted on this pest on pepper crops in Egypt.

The objective of the present study was to evaluate the reduction of *F. occidentalis* populations on pepper crop under greenhouse conditions by releasing the predatory phytoseiid mites in combination of using of blue sticky traps and/or by applying pesticides.

Materials and methods

The stored grain mite, Tyrophagous putrescentiae (Schrank) (Acari: Acaridae), was collected from debris samples of wheat bran, using modified Tullgren funnels (Krantz and Walter 2009). T. putrescentiae was reared on a mixture of wheat bran and bakery dry yeast at 25 °C in top vented plastic containers (6 × 3 cm, diameters × depth). Mass rearing of the phytoseiid predatory mites was conducted on Tyrophagous putrescentiae. Tested phytoseiid mites were reared constantly on a mixture of all stages of the acarid mite under controlled conditions of 25 ± 2 °C in top-vented plastic containers (6 cm diameter and 10 cm depth). All units were kept in large boxes provided with NaCl-saturated solution for keeping up RH inside the boxes as 75% (Winston and Bates 1960). Releasing of phytoseiid mites was conducted by using of a carrier which consisted of a mixture of wheat bran and vermiculate material by 1:1 ratio.

Greenhouse experiments

Two field experiments were conducted in two symmetrical pepper greenhouses (6 m width \times 45 m length \times 2.5 m height) in a horticulture farm at Qaha district, Qalyobia Governorate, Egypt, during the period from 12 May to 7 July throughout two successive seasons, 2017 and 2018, under natural infestation conditions. The two greenhouses were cultivated by pepper (Top Star var.). Recommended agricultural practices were applied.

The obtained *F. occidentalis* individuals from the infested greenhouses were preserved in vials containing 70% ethanol alcohol until identification took place at the Taxonomy Research Department, Plant Protection Research Institute (PPRI), Agriculture Research Center (ARC), Giza, Egypt.

The first experiment was carried out to determine the efficiency of four predatory phytoseiid mites (in the presence of blue sticky trap) to reduce F. occidentalis population in pepper greenhouse. The greenhouse was divided into five blocks (9 \times 6 m²/block and 9 \times 2 m²/ replicates), and each was divided to three replicates. Each block was completely separated by acrylic sheet to prevent mite movement. Thirty blue sticky traps were utilized for attracting of F. occidentalis adults during both seasons in all blocks (each block treatment contained 6 traps). Sticky sheet was (18 cm width × 28 cm length) with a sticky material exposed on both sides. The blue sticky sheet was weekly replaced by new ones. The blue sheet trap (blue sensitive plastic sheet) was obtained from PPRI. The rates and times of releasing of the four predatory phytoseiid mites were presented in Table 1; the releasing of four predatory mites was conducted throughout the flowering stage of pepper plant, due to the F. occidentalis infestation level was highly population.

The second experiment was conducted in a neighbor pepper greenhouse, divided as the first one. All management control programs of WFT used spray of chemical, to be compared with untreated block (Table 2). Comparison between the four programs of insecticides, against WFT and untreated control (check), was conducted throughout the two successive seasons, 2017 and 2018. Pesticide applications were conducted using calibrated handheld compression sprayer (Kwazar) before application. Each block treatment was completely separated by a plastic sheet to avoid pesticide drift.

The mean numbers of WFT individuals (adults and nymphs/10 pepper flowers) were recorded weekly till the end of the experiments. Also, the mean numbers of WFT adults attracted per blue sheet were recorded. Inspection of WFT per ten pepper flowers was carried out, using a white plan sheet (23 cm width \times 33 cm length) as inspection plate. The plate was sprayed by water to avoid insects escaping.

Data analysis

Statistical analyses were performed, using SAS program computer including *F*-test (SAS Institute 2003). Revised least significant differences (LSD) at 5% level of probability were used for comparing means.

Results and discussion

In the untreated check (control), the WFT population recorded three peaks on 19 May, 9 June, and 30 June, 2017, by 33.67, 255.33, and 185.67 individuals per 10 flowers, respectively (Table 3). In the treatments, after 2 June 2017 (first date of releasing), the *F. occidentalis* individuals decreased by releasing the phytoseiid mites. In the treatment of *N. barkeri* and *N. arundonaxi*, a low

Table 1 Releasing rates and dates of tested phytoseiid species against *Frankliniella occidentalis* on pepper plants during the two seasons of 2017 and 2018

Phytoseiid species	Plant from which collected	Rate of application	2017		2018	
			Date of first releasing	Date of second releasing	Date of first releasing	Date of second releasing
Cydnoseius negevi (Swirski and Amitai)	Cynodon dactylon (L.) Pers. (Family: Poaceae)	400–500 individuals/plant	2 June	30 June	19 May	9 June
<i>Neoseiulus arundonaxi</i> (Metwally and Sanad)	<i>Arundo donax</i> L. (Family: Poaceae)					
Neoseiulus barkeri (Hughes)	Cynodon dactylon (L.) Pers. (Family: Poaceae)					
Neoseiulus bicaudus (Wainstein)	<i>Arundo dona</i> x L. (Family: Poaceae)					

population of WFT (24.33 and 62.67 individuals/10 flowers, respectively, in season 2017) was recorded after 1 week from releasing. At the second releasing of the four tested predatory mites, the population of WFT increased on the block with *N. bicaudus* and *C. negevi* (176.67 and 163.73 individuals/10 flowers, respectively, in 2017 season) (Table 3).

After the second releasing (30 June 2017), the WFT population was sharply dropped after 2 weeks from releasing all phytoseiid mites. The total mean numbers

of WFT were 47.60, 39.86, 67.25, 73.87, and 81.13 individuals per 10 flowers in the case of *N. barkeri*, *N. arundonaxi*, *N. bicaudus*, *C. negevi*, and untreated check, respectively, during the 2017 season (Table 3). Generally, the releasing of both *N. barkeri* and *N. arundonaxi* was more effective for WFT suppression in the pepper greenhouse.

In the second season, 2018, the same trend was reported in the four releases of phytoseiid mites against WFT infestation on pepper plants under greenhouse

Table 2 Application of four programs for *Frankliniella occidentalis* management on pepper plants during the two seasons of 2017 and 2018

Programs	Treatments	Applications time	Application	
		2017	2018	rate/100 L
1	Sequentially spraying of	Start on 19 May at 15 days interval	Start on 15 May at 15 days interval	
	Sulfoxaflor 24% SC	(4 applications)	(4 applications)	50 mL
	Imidacloprid 35% SC			75 mL
	Azadirachtin 0.18% EC			200 mL
	Mineral oil 98% EC			1.5 L
2	Sequentially spraying of			
	Amizatine 20% EC			375 mL
	Acetamiprid 20% SP			25 g
	Azadirachtin 0.18% EC			200 mL
	Mineral oil 98% EC			1.5 L
3	Sequentially spraying of			
	Chlorfenapyr 24% SC			60 mL
	Imidacloprid 70% WDG			60 g
	Azadirachtin 0.18% EC			200 mL
	Agricultural potassium soap 49% liquid			2 L
4	Sequentially spraying of			
	Azadirachtin 0.18% EC			200 mL
	Mineral oil 98% EC			1.5 L
	Agricultural potassium soap 49% liquid			2 L
	Agricultural potassium soap 49% liquid			2 L
Check (control)	Untreated pepper plant	-	-	-

Table 3 Mean number of *Frankliniella occidentalis* adult and nymphs after the releasing of the four phytoseiid mites at tenth week after plantation on pepper greenhouse during the two seasons of 2017 and 2018

Inspection dates	Mean numbers of F. occidantalis adults and nymphs/ 10 flowers										
	2017					2018					
	Control	Neoseiulus barkari	Neoseiulus arundonaxi	Neoseiulus bicaudusNeoseiulus bicaudus	Cydnoseius negevi	Control	Neoseiulus barkari	Neoseiulus arundonaxi	Neoseiulus bicaudusNeoseiulus bicaudus	Cydnoseius negevi	
May, 12 th	16.40	12.83	18.77	20.67	31.70	92.10	63.30	96.20	72.37	61.27	
19 th	33.67	14.67	15.33	31.67	26.67	96.03	85.07	66.27	158.20	110.07	
26 th	11.33	17.47	19.67	14.67	10.33	79.10	39.37	53.20	61.97	63.93	
June, 02 nd	115.50	166.67	100.33	171.33	149.67	84.57	89.93	23.90	67.87	96.13	
09 th	255.33	24.33	62.67	101.67	83.67	102.40	95.72	61.93	74.15	96.75	
16 th	66.33	84.33	24.67	161.67	129.33	68.57	39.40	11.07	26.25	10.76	
23 th	103.33	45.33	32.37	31.33	35.33	93.73	23.10	25.78	17.01	16.01	
30 th	185.67	70.37	70.33	176.67	163.73	153.01	4.01	10.03	15.00	6.01	
July, 7 th	16.67	37.00	36.33	19.00	32.00	110.02	2.02	7.03	5.00	4.00	
14 th	7.02	3.00	18.17	10.00	10.07	94.01	6.02	2.03	2.00	1.00	
Mean	81.13 ^a	47.60 ^{bc}	39.86 ^c	73.87 ^a	67.25 ^{ab}	97.35 ^a	44.79 ^b	35.74 ^b	49.98 ^b	46.59 ^b	
± SE	±8.81	± 10.61	± 5.88	± 8.56	± 7.86	± 4.57	± 7.05	± 14.96	± 12.68	± 5.18	
F value	6.18					12.31					
LSD	22.248						19.366				

Values in row followed by similar letter were not significant at 5% level of probability

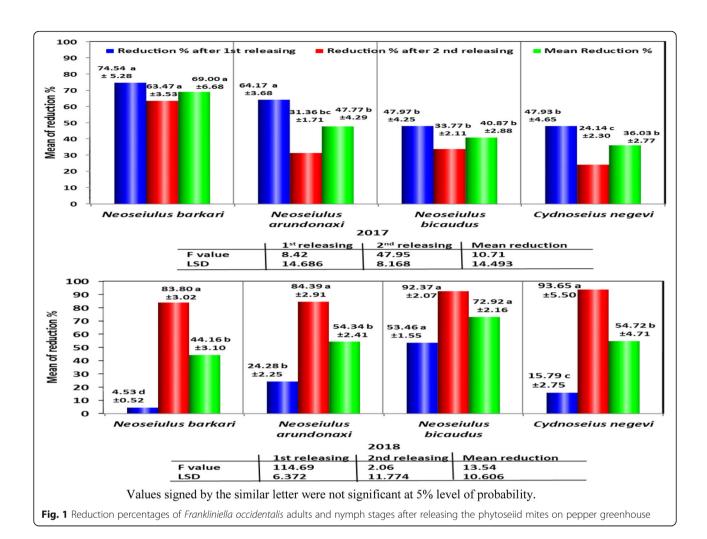
conditions. The releasing of these predators was applied on 19 May and 9 June 2018. During the 2018 season, the population of WFT-infested pepper plant was slowly decreased after the first releasing of predators, while the WFT suppression was rapidly dropped after the two releases in the 2018 season (Table 3). Insignificant differences were recorded among the four releases of the phytoseiid mites throughout the season 2018. In general, the block, released by N. barkeri and N. arundonaxi, harbored the low infestation of WFT, F. occidentalis, during the two successive seasons, 2017 and 2018, under greenhouse. A higher reduction percentage rate in population of WFT on pepper was detected when N. barkeri was released in 2017 (74.54, 63.47, and 69.00% reduction after first and second releases and mean reduction after two releases, respectively), followed by N. arundonaxi recording 64.17, 31.36, and 47.77% reduction in the case of first and second releases and overall mean reduction after two releases, respectively (Fig. 1). However, N. bicaudus and C. negevi recorded the low reduction percentages (overall means of 40.87 and 36.0% reduction, respectively) on *F. occidentalis* on pepper plant under greenhouse conditions in the 2017 season (Fig. 1).

On the other hand, *N. bicaudus* and *C. negevi* caused the higher reduction of *F. occidentalis* individuals after two releasing processes than the other two predators, *N. barkeri* and *N. arundonaxi*, in season 2018. The reduction percentage was 53.46, 24.28, 15.79, and 4.53% after first

releases of *N. bicaudus*, *N. arundonaxi*, *C. negevi*, and *N. barkeri*, respectively, in the second season 2018. After the second releasing, the reduction was 93.65, 92.37, 84.39, and 83.80% for *C. negevi*, *N. bicaudus*, *N. arundonaxi*, and *N. barkeri* in 2018, respectively. The overall mean reduction percentage was 72.92, 54.72, 54.34, and 44.16% in the second season 2018 with *N. bicaudus*, *C. negevi*, *N. arundonaxi*, and *N. barkeri*, respectively (Fig. 1).

Generally, the overall mean reduction percentage can be arranged as the following: 56.89, 56.58, 51.05, and 45.38% with *N. bicaudus*, *N. barkeri*, *N. arundonaxi*, and *C. negevi*, in seasons 2017 and 2018, respectively.

Data in Table 4 show the combined effect of releasing tested phytoseiids and the used of blue sticky trap on pepper plant under greenhouse conditions. The population of WFT individuals/blue sticky sheet recorded three peaks through the tested seasons, 2017 and 2018. The combined effect of predators and sticky traps on the WFT infestations decreased sharply on pepper plants than in the untreated pepper plants (check) (Table 4). The use of predators and blue sticky traps together was more effective for thrips suppression. The means of F. occidentalis individuals/blue sheet during 2017 and 2018 seasons are shown in Table 4. The overall mean of WFT individuals/blue sticky sheet was the highest (63.80 and 34.10 individuals/blue sheet) in block of untreated check, followed by the block of treated one with N. arundonaxi releasing and blue sticky traps (37.90 and 31.20 individuals/blue sticky traps) throughout seasons 2017 and



2018, respectively. Subsequently, the predator-sticky trap combination was more appropriate for WFT controlling on pepper plant under greenhouse conditions through using *N. barkeri*, *N. bicaudus*, and *N. arundonaxi*.

For successful management of *F. occidentalis* on pepper plants under greenhouse conditions, determination of the thrips damages was critical because the pest had a broad resistance range against insecticides for different chemical categories (Demirozer et al. 2012 and Mouden et al. 2017).

In the present study, using of blue sticky traps was more suitable for reducing and attracting of WFT individuals. Broughton and Harrison (2012) reported that *F. occidentalis* and *T. tabaci* were attracted to blue, yellow, and white sticky traps than other tested traps, red, black, and green sticky traps, also in which the blue sticky traps caught the thrip individuals more than the yellow traps. However, current recommendations for attracting of thrips were yellow sticky traps and/or flowers (Hardy et al. 2005). Introducing both blue and yellow sticky cards above the plant was the main method for scouting

and controlling of thrips by used about 10-40 sticky cards (Cloyd 2009).

The obtained data reported that the mean reduction percentage of F. occidentalis infestations could be arranged as 56.89, 56.58, 51.05, and 45.38% with N. bicaudus, N. barkeri, N. arundonaxi, and C. negevi, respectively, during the seasons 2017 and 2018. These results were nearly comparable with those findings by Hessien and Parrella (1990) who stated that releasing of N. barkeri led to reduce F. occidentalis individual up to 41% and releasing N. cucmeris or N. barkeri was successful for suppressing of thrips in flower crops. The releasing process was conducted per plant as nearly similar to that reported by Hessien and Parrella (1990) who released N. cucmeris and N. barkeri at (2.5 mites/leaf) against the WFT population. However, the better WFT suppression was achieved when N. cucumeris released at 106 individuals/m²/ biweekly (Van Driesche et al. 2006), while weekly releasing of N. cucumeris (180 individuals/m²) was suitable for F. occidentalis controlling (Bennison et al. 2002). For intensification biological control, N. bicaudus, N. barkeri, and N.

Table 4 Mean numbers of WFT, *Frankliniella occidentalis*, adults/blue sticky trap after the releasing of phytoseiid mites on pepper greenhouse

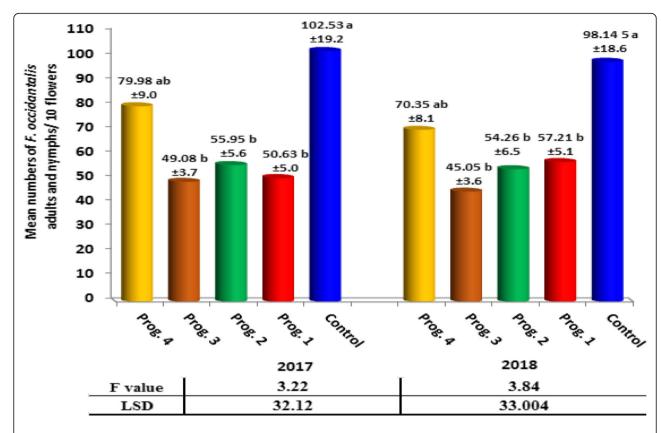
Inspection dates	Mean numbers of F. occidantalis adults / Blue sheet										
	2017					2018					
	Control	Neoseiulus barkari	Neoseiulus arundonaxi	Neoseiulus bicaudusNeoseiulus bicaudus	Cydnoseius negevi	Control	Neoseiulus barkari	Neoseiulus arundonaxi	Neoseiulus bicaudusNeoseiulus bicaudus	Cydnoseius negevi	
May, 12 th	35.80	24.90	81.40	41.90	19.00	28.80	36.00	34.40	32.00	25.80	
19 th	37.80	52.70	45.80	29.30	29.00	24.20	25.60	41.00	19.80	23.20	
26 th	137.50	24.00	34.00	97.80	53.00	20.80	19.40	19.80	16.20	19.40	
June, 02 nd	143.40	13.10	22.20	101.60	56.00	9.60	6.00	28.80	18.60	10.20	
09 th	23.20	34.10	6.80	10.10	18.40	67.80	14.00	36.80	26.60	18.20	
16 th	108.70	7.40	42.00	25.50	47.50	77.20	53.20	57.40	57.00	50.20	
23 th	54.90	27.50	29.50	43.00	21.00	66.40	28.80	22.00	14.20	21.20	
30 th	29.50	1.20	2.30	19.40	4.10	36.60	39.60	45.80	42.00	36.80	
July, 7 th	46.40	36.30	10.30	10.10	19.90	6.60	12.40	15.40	10.60	13.60	
14 th	21.00	0.20	1.30	0.00	1.00	3.20	8.20	10.40	14.60	4.60	
Mean	63.80 ^a	22.10 ^b	27.60 ^b	37.90 ^b	26.90 ^b	34.10 ^a	24.30 ^b	31.20 ^{ab}	25.20 ^{ab}	22.30 ^b	
± SE	± 8.98	± 3.22	± 4.60	± 6.38	± 3.61	± 4.88	± 3.06	± 3.00	± 2.79	± 2.64	
F value	8.47				2.19						
LSD	16.098						9.431				

Means followed by the same letter were not significant at 5% level of probability

arundonaxi framework was an augmentative progress against WFT infestation on pepper greenhouse.

Fortunately, pepper greenhouse provides the suitable media for WFT development and infestation. The chemical control of this pest was difficult. According to these programs, the combined of chemical pesticide and biopesticide altogether was used in all these programs of WFT control. The mean numbers of F. occidentalis recorded a high value on check pepper block (untreated check) (89.14 individuals/10 flowers), followed by the fourth program (70.35 individuals/10 flowers) during the first season 2017, while the other three programs stated insignificant differences among them during 2017 season (57.21, 54.26, and 45.05 individuals/10 flowers for the first, second, and third programs, respectively). Similarly, the same trend of results was obtained during the second season 2018. The fourth program of pesticides was reported as a high value of WFT individuals/10 flowers, but the low value was observed on each of third, first, and second programs (49.08, 50.63, and 55.95 WFT individuals/ 10 flowers during season 2018, respectively (Fig. 2)). Finally, the best WFT control management was the third program, in which sequentially spraying of chlorfenapyr, imidacloprid, azadirachtin, and agricultural potassium soap was used. Data in Fig. 2 illustrates a significant difference between the combination of chemical pesticides and biopesticides in the four tested programs against WFT on pepper plant under greenhouse conditions.

The present data was in harmony with those findings by Hanafy (2015), in which acetamiprid and thiamethoxam gave a significantly high suppression of F. occidentalis, followed by carbosulfan, spinosad, and spinetoram after 14 days from applications. WFT management key by insecticides can be initiated at the low population of F. occidentalis to avoid overlapping of this pest; once it reaches a high population, it is very difficult to control (Cloyd 2009). In the present study, nine different insecticides in four programs were tested against adults and nymphs stages, because the most currently recommended insecticides only suppressed or killed both thrips stages, with no activity on both the eggs and pupal stages (Seaton et al. 1997). The primary rule to prevent or reduce the WFT resistance from insecticide was rotating insecticides use (Robb and Parrella 1995). In the present study, numerous pesticides were rotated against WFT population on pepper plants under greenhouse conditions. For example, WFT populations were recorded to be resistance against carbamate, pyrethroid, and organophosphates (Kontsedalov et al. 1998). Finally, releasing of N. bicaudus and N. barkeri in combination with blue sticky cards reduced the WFT by 56.89 and 56.58%, respectively. The application of the third program (sequential spraying of chlorfenapyr, imidacloprid, azadirachtin, and agricultural potassium soap) gave the best results in reducing the (WFT) (47.1 mean numbers of individuals/10 flowers during the two seasons).



Means followed by the same letter were not significant at 5% level of probability.

Fig. 2 Implement of Frankliniella occidentalis managing by sequentially four insecticide programs in pepper greenhouse

Conclusion

In the present study, releasing of *N. bicaudus*, *N. barkeri*, *N. arundonaxi*, and *C. negevi* in combination with the blue sticky traps reduced *F. occidentalis* individuals up to 56% and can be recommended for controlling *F. occidentalis* on pepper plantations in the greenhouses.

Acknowledgements

Not applicable

Authors' contributions

ASS and GMH contributed to the suggesting and putting of the idea, preparing of the manuscript writing and finishing of the paper, and data analysis. All authors contribute by 100% participation. Both authors read and approved the final manuscript.

Funding

No funding

Availability of data and materials

Not applicable

Ethics approval and consent to participate

Not applicable

Consent for publication

Not applicable

Competing interests

The authors declare that they have no competing interests.

Author details

¹Vegetable and Aromatic plant Mites Department, Plant Protection Research Institute, Agricultural Research Center, 12618 Dokii, Giza, Egypt. ²Vegetable, Medicinal, Aromatic and Ornamental Plant Pests Department, Plant Protection Research Institute, Agricultural Research Center, Giza, Egypt.

Received: 4 August 2019 Accepted: 29 October 2019 Published online: 30 December 2019

References

Arthurs S, Mckenzie CL, Chen J, Dogramaci M, Brennan M, Houben K, Osborne L (2009) Evaluation of *Neoseiulus cucumeris* and *Amblyseius swirskii* (Acari: Phytoseiidae) as biological control agents of chilli thrips, *Scirtothrips dorsalis* (Thysanoptera: Thripidae) on pepper. Biol Cont 49(1):91–96

Bennison J, Maulden K, Maher H (2002) Choice of predatory mites for biological control of ground-dwelling stages of western flower thrips within a 'push-pull' strategy on pot chrysanthemum. Bulletin OILB/SROP 25:9–12

Bethke, J.; Chamberlin, J.; Dobbs, J.; Faver, M.; Heinz, K.; Lindquist, R.; Ludwig, S.; Mckenzie, C.L.; Murphy, G.; Oetting, R.; Osborne, L.; Palmer, C.; Parrella, M.; Rechcigl, N. and Yates, R. (2010): Thrips management program for plants for planting. Mid Florida Research & Education Center, Univ. Florida, 1-9 pp.

Broughton S, Harrison J (2012) Evaluation of monitoring methods for thrips and the effect of trap colour and semiochemicals on sticky trap capture of thrips (Thysanoptera) and beneficial insects (Syrphidae, Hemerobiidae) in deciduous fruit trees in Western Australia. Crop Protection 42:156–163

- Cloyd RA (2009) Western flower thrips (*Frankliniella occidentalis*) management on ornamental crops grown in greenhouses: have we reached an impasse? Pest Technology 3:1–9
- Demirozer O, Tyler-Julian K, Funderburk J, Leppla N, Reitz SR (2012) Frankliniella occidentalis (Pergande) integrated pest management programs for fruiting vegetables in Florida. Pest Manag Sci 68:1537–1545
- Dogramaci M, Arthurs SP, Chen J, McKenzie CL, Irrizary F, Osborne LS (2011) Management of Chilli thrips, *Scirtothrips dorsalis* (Thysanoptera: Thripidae) on peppers by *Amblyseius swirskii* (Acari: Phytoseiidae) and Orius insidiosus (Hemiptera: Anthocoridae). Biol Cont. 59(3):340–347
- Hanafy ARI (2015) Population fluctuation, host preference, damage and control of western flower thrips, *Frankliniella occidentalis* (Pergande) on some vegetable crops in Egypt. Egypt Acad J Biolog Sci 8(1):145–156
- Hardy, S.; Steiner, M.; Thwaite, G. (2005): Plague thrips and western flower thrips. In:
 Integrated Pest and Disease Management for Australian Summerfruit. NSW
 Department of Primary Industries. NSW. Summerfruit Australia Inc. 65-69 pp.
- Hessein NA, Parrella MP (1990) Predatory mites help control thrips on floriculture crops. California Agriculture 44:19–21
- Immaraju JA, Paine TD, Bethke JA, Robb KL, Newman JP (1992) Western flower thrips (Thysanoptera: Thripidae) resistance to insecticides in coastal California greenhouses. J Econ Entomol 85:9–14
- Kirişik M, Erler F (2017) The usage possibilities of entomopathogenic fungi in the control of western flower thrips, *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae). Turkish J Entomol 7(4):293–303
- Knapp M, van Houten Y, Hoggerbrugge H, Bolckmans K (2013) Amblydromalus limonicus (Acari: Phytoseiidae) as a biocontrol agent: review and new findings. Acaralogia 53:102–202
- Kontsedalov S, Weintraub PG, Horowitz AR, Ishaaya I (1998) Effects of Insecticides on Immature and Adult Western Flower Thrips (Thysanoptera: Thripidae) in Israel. J Econ Entomol 91(5):1067–1071
- Krantz GW, Walter DE (2009) A manual of acarology: third edition, Texas Tech. University Press, USA, 816 pp
- Messelink GJ, Steenpaal SEF, Ramakers PMJ (2006) Evaluation of phytoseiid predators for control of Western flower thrips on greenhouse cucumber. Biocontrol 51:753–768
- Minakuchi C, Inano Y, Shi X, Song D, Zhang Y, Miura K, Sonoda S (2013)

 Neonicotinoid resistance and cDNA sequences of nicotinic acetylcholine receptor subunits of the western flower thrips *Frankliniella occidentalis* (Thysanoptera: Thripidae). Appl Entomol Zool 48(4):507–513
- Mouden S, Sarmiento KF, Klinkhamer PGL, Leiss KA (2017) Integrated pest management in western flower thrips: past, present and future. Pest Manag Sci 73:813–822
- Ramakers PMJ (1988) Population dynamics of the thrips predators *Amblyseius* mckenziei and *Amblyseius cucumeris* (Acarina: Phytoseiidae) on sweet pepper. Netherlands J Agricul Sci 36:247–252
- Robb KL, Newman J, Virzi JK, Parrella MP (1995) Insecticide resistance in western flower thrips. In: Parker BL, Skinner M, Lewis T (eds) Thrips Biology and Management. Plenum Press, New York, pp 341–346
- Robb KL, Parrella MP (1995) IPM of western flower thrips. In: Parker BL, Skinner M, Lewis T (eds) Thrips Biology and Management. Plenum Press, New York, pp 365–370
- SAS Institute (2003): Statistical Analysis System, SAS Release 9.1 for windows, SAS Institute Inc.Cary, NC, USA.
- Seaton KA, Cook DF, Hardie DC (1997) The effectiveness of a range of insecticides against western flower thrips (*Frankliniella occidentalis*) (Thysanoptera:Thripidae) on cut flowers. Aust J Agric Res 48:781–788
- Shalaby, H.H. (2015): Preliminary study on the control of western flower thrips, Frankliniella occidentalis (Pergande) in pepper crop greenhouses in Qalyubia Governorate, Egypt. J Plant Prot Path., Mansoura Univ., 6(1): 155-167.
- Van Driesche RG, Lyon S, Stanek EJ, Xu B, Nunn C (2006) Evaluation of efficacy of Neoseiulus cucumeris for control of Western flower thrips in spring bedding crops. Biol Control 36:203–215
- Winston PW, Bates DH (1960) Saturated solutions for the control of humidity in biological research. Ecology 41:232–237

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Submit your manuscript to a SpringerOpen journal and benefit from:

- ► Convenient online submission
- ► Rigorous peer review
- ► Open access: articles freely available online
- ► High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ▶ springeropen.com