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# Can *Beauveria bassiana* reduce the root lesion nematode, *Pratylenchus thornei*, infection on wheat?

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## Abstract

**Background:** Limited number of control methods against root-lesion nematodes has increased the search for alternative management strategies. Due to secondary metabolites such as toxins and enzymes secreted by the entomopathogenic fungus, *Beauveria bassiana*, the number of studies on nematicidal potentials on many plant parasitic nematodes has increased. Almost no work was done on the nematicidal activity of *B. bassiana*, which is widely used as a bio-insecticide, on the root lesion nematode, *Pratylenchus thornei*, commonly found on wheat in Turkey. In the present study, it was aimed to determine the pathogenicity of two native *B. bassiana* isolates (BIM-001 and BY2) obtained from Turkey against *P. thornei* on wheat under controlled conditions.

**Results:** Spore suspension of *B. bassiana* isolates affected *P. thornei* soil, root, final nematode density and reproduction rate at different degrees according to spore concentration. The soil, root, total nematode density, and reproduction rate in the control on İközce cv. were 1234.0 adult + larvae/500 g soil, 1105.0 adult + larvae/per pot, 2341.0 (soil + root density) and 5.8 (PF(final nematode density/PI (initial inoculum density))), respectively. It was determined that these parameter values were at the lowest at  $10^8$  spore/ml than other concentrations in both isolates of *B. bassiana*. In  $10^8$  spore/ml concentrations of BIM-001 isolate, while *P. thornei* soil, root, total nematode density, and reproduction rate on İközce were 641.7 adult + larvae/500 g soil, 930.9 adult + larvae/per pot, 1572.6 (soil + root density) and 3.9 (PF/PI), respectively, these parameter values were found to be 645.2 adult + larvae/500 g soil, 849.0 adult + larvae/per pot, 1492.2 (soil + root density) and 3.6 (PF/PI) at  $10^8$  spore/ml concentrations of BY2 isolate. It was observed that the reproduction rate of *P. thornei* decreased at  $10^8$  spore/ml concentration of BIM-001 and BY2 isolates compared to the control.

**Conclusion:** Native *B. bassiana* isolates of BIM-001 and BY2 reduced *P. thornei* on wheat and the concentration was important for the pathogenicity of *B. bassiana*. While the reproduction rate of *P. thornei* at  $10^8$  and  $10^7$  spores/ml concentrations in both isolates decreased than the control. At  $10^6$  spore/ml concentration it was found similar to the control. Even though *B. bassiana* is known as an effective biocontrol agent against insects and some plant-parasitic nematodes, more detailed studies should be done on its effect on *P. thornei*.

**Keywords:** *Beauveria bassiana*, Wheat, *Pratylenchus thornei*, Root lesion nematode, Virulence

## Background

The root lesion nematode, *Pratylenchus thornei* (Sher et Allen) (Tylenchida: Pratylenchidae), is known as a parasite of the roots of cereals and legumes in many countries around the world causing economic losses (Castillo and Vovlas 2007; Thompson et al. 2021). This nematode species is commonly found in cereal production areas

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in Turkey (Göze Özdemir et al. 2021). This nematode is polycyclic and completes its life cycle in about 45 days under optimum conditions (Larson 1959). Therefore, population densities increase exponentially during the production season (Thompson 2015). While *P. thornei* feeds and multiplies in the root cortex, it also disrupts the function of the root. It causes characteristic lesions that inhibit the plant's ability to absorb water and nutrients, and cause yield loss in susceptible crops (Tan et al. 2013). In addition, these lesion areas constitute areas of infection for many fungal agents (Mallaiah et al. 2014). It is suggested that root lesion nematodes can be controlled with methods such as ensuring field hygiene, cultivation resistant or tolerant wheat varieties, or rotation with non-host crops (Owen et al. 2010). The absence of licensed nematicides for the control of nematodes in cereals also limits the possibilities of controlling this pest group and increases the search for alternative and sustainable control methods (Thompson et al. 2021). As a result of these searches, it has been reported that studies on the effects of different fungi on plant-parasitic nematodes have increased due to the ability to derive compounds from toxins, enzymes, or metabolites (Göze Özdemir and Arıcı 2021).

*Beauveria bassiana* (Balsamo.-Criv.) Vuillemin 1912 (Hypocreales/Cordycipitaceae) is a widely known entomopathogenic fungus worldwide and there are many isolates commercially used for the control of various arthropod species (Mwaura et al. 2017). *Beauveria*, besides insects, also produces organic compounds that affect nematode behavior (Hummadi et al. 2021). These compounds offer the potential to be used as fumigants to eliminate the free-living stages of plant-parasitic nematodes (Khoja et al. 2021). The mechanism of antagonism of *B. bassiana* includes antibiosis, competition and induced systemic resistance (Devi 2019). Basically, chitinases, lipases, and proteases are the most important of all the enzymes produced by *B. bassiana* (Amobonye et al. 2020). However, in different studies, it has been determined that it has the ability to produce other enzymes such as amylase, asparaginase, cellulase, galactosidase (Petlamul and Boukaew 2019). In different studies, the presence of beauvericin, bassianolide, bassiacridin, and oosporein toxins in *B. bassiana* culture supernatants have been reported (Ortiz-Urquiza et al. 2010). There are different studies showing that fermented filtrate products from *B. bassiana* have potential in the control of important plant-parasitic nematodes such as *Ditylenchus destructor* Thorne, 1945, *Meloidogyne hapla* Chitwood, 1949, *M. incognita* (Kofoid and White 1919) Chitwood, 1949, *Heterodera avenae* (Wollenweber 1924) and *Pratylenchus* sp. (Youssef et al. 2020).

It is known that the use of resistant-tolerant varieties has priority in the control of root lesion nematodes in wheat worldwide and a different alternative method has not been recommended yet. The lack of any detailed study on the effect of *B. bassiana*, which is widely used as a bio-insecticide in pest control, on root lesion nematodes led to the planning of this study. This study aimed to reveal the pathogenicity of two native *B. bassiana* isolates obtained from Turkey against *P. thornei* in wheat under controlled conditions.

## Materials and methods

### Materials

The study was carried out with the SK11 isolate of *P. thornei* which was obtained from the wheat production area of Şarkikaraağaç district (Turkey) in Isparta Province (Göze Özdemir et al. 2021) and whose mass production in the laboratory continued according to Zuckerman et al. (1985)'s carrot disk method. BIM-001 of *B. bassiana* isolated from *Leptinotarsa decemlineata* Say (Coleoptera: Chrysomelidae) in potato fields in Isparta (Sarı, 2020) and BY2 isolate isolated from *Haplothrips* sp. in Burdur wheat fields were used in the study (Uzun, 2020). The wheat material of the study was İkişze cultivar, which was found to be susceptible to root lesion nematode in previous studies conducted in Isparta Province (Sögüt et al. 2011).

### Preparation of nematode inoculum

After mass production of *P. thornei*, carrot disks in 9 cm diameter Petri dishes were cut into small pieces, sterile water was added to cover the Petri dish. After two days, the nematodes that got into the water were taken with a tape measure and the precipitation process was carried out. The obtained nematode suspension was counted using a binocular microscope and approximately 400 *P. thornei* (adult + larvae) were transferred into Eppendorf tubes with 1 ml of sterile water (Kepenekçi et al. 2018).

### Preparation of the fungal inoculum

*Beauveria bassiana* BIM-001 and BY2 isolates were cultured on Potato Dextrose Agar (Sigma-Aldrich, Germany) medium in 10 cm diameter Petri dishes and incubated at 25 °C for 10 days in dark conditions. After ten days, distilled water was poured onto the fungus culture dishes and the dishes were rubbed with a spreader. Then, a filter was placed on the beaker and distilled water + spores were poured onto the filter. Spore suspension concentrations  $10^6$ ,  $10^7$ ,  $10^8$  spores/ml were prepared

by counting the spores on the thoma slide (Youssef et al. 2020).

#### Determination of pathogenicity of *Beauveria bassiana* against *Pratylenchus thornei* on wheat

The study was carried out in 500 cc plastic pots with 500 g of autoclaved soil mixture (68% sand, 21% silt, and 11% clay) in a climate room with  $25 \pm 2$  °C and  $60 \pm 5\%$  proportional humidity under controlled conditions. Three wheat plants in pot were considered as one replica. Experiments were set up separately for *B. bassiana* BIM-001 and BY2 isolate. The study was carried out with 10 replications of both entomopathogenic fungus isolates according to a randomized plot design for each suspension concentration ( $1 \times 10^6$ ,  $1 \times 10^7$ ,  $1 \times 10^8$  spores/ml). One week after sowing the seeds, 400 *P. thornei* individuals (larvae + adult) were inoculated in each pot with the help of plastic pipettes into the holes drilled to 2–3 cm soil depth around the root zone (Kepenekçi et al. 2018). Five ml of the spore suspensions were prepared immediately after the nematode inoculation was applied to each pot with the help of a measuring tape. Only wheat plants treated with *P. thornei* were used as control. Experiments were terminated after 9 weeks. At the end of this period, plant and root length (cm) and fresh weight (g) of wheat plants were recorded. *P. thornei* individuals were obtained from soil and root using the Baermann funnel method (Hooper et al. 2005). Then, nematodes counts were made under a binocular microscope at 10X magnification, and after the total nematode density was obtained, the reproduction rates were calculated with the formula; Reproduction rates = Final nematode density (PF)/number of inoculated nematodes (PI).

#### Statistical analysis

SPSS (version 20.0) program was used for the statistical analysis of the data obtained in the study and analysis of variance (ANOVA) was performed to test the differences between the means. “Tukey” test was used when variances were homogeneous to determine the means of different groups.

#### Results

In the study, the mean of soil nematode density was lower than the control plants that were only treated with *P. thornei* in all concentrations of *B. bassiana* BIM-001 isolate. However, it was determined that there was non-significant difference between the concentrations of BIM-001 isolate in terms of mean of soil nematode density ( $P \geq 0.05$ ). The mean root nematode density of  $10^8$  ( $930.9 \pm 45.9$ /per pot) and  $10^7$  ( $960.0 \pm 68.7$ /per pot) spores/ml concentrations of BIM-001 isolate was lower than the control ( $1105.0 \pm 111.2$ /per pot), but the difference between them was non-significant ( $P \geq 0.05$ ). The highest mean of total nematode density and reproduction rate was determined at  $10^6$  spore/ml concentration of BIM-001 isolate and control. Although the mean of total nematode density and reproduction rate parameters of  $10^8$  spore/ml concentrations of BIM-001 isolate were lower than  $10^7$  spore/ml concentrations, non-significant difference was found between them ( $P \geq 0.05$ ). However, it was determined that they were lower than the control (Table 1).

The mean soil nematode density of  $10^8$ ,  $10^7$ , and  $10^6$  spores/ml concentrations of *B. bassiana* BY2 isolate was lower than the control. It was determined that the lowest mean of *P. thornei* soil, root, total nematode density and reproduction rate was found at  $10^8$  spore/ml

**Table 1** Effect of *Beauveria bassiana* BIM-001 and BY2 isolates on soil, root, total nematode density and reproduction rate of *Pratylenchus thornei*

Parameters					
Isolates	Spore Concentration	Soil nematode density (500 g soil)	Root nematode density (per pot)	Final nematode density (soil + root)	Reproduction rate (PF/PI)
<i>Mean ± Standard error*</i>					
BIM-001	$10^8$	$641.7 \pm 27.1^b$	$930.9 \pm 45.9^b$	$1572.6 \pm 36.2^b$	$3.9 \pm 0.0^b$
	$10^7$	$717.4 \pm 43.0^b$	$960.0 \pm 68.7^b$	$1819.0 \pm 65.6^b$	$4.5 \pm 0.1^b$
	$10^6$	$859.0 \pm 76.9^b$	$1429.6 \pm 77.1^a$	$2147.0 \pm 109.2^a$	$5.3 \pm 0.2^a$
	Control	$1234.0 \pm 71.6^a$	$1105.0 \pm 111.2^b$	$2341.0 \pm 53.3^a$	$5.8 \pm 0.1^a$
BY2	$10^8$	$645.2 \pm 35.0^c$	$849.0 \pm 97.7^B$	$1492.2 \pm 99.7^C$	$3.6 \pm 0.2^C$
	$10^7$	$769.6 \pm 36.3^B$	$908.0 \pm 111.3^{AB}$	$1809.8 \pm 92.5^B$	$4.4 \pm 0.2^B$
	$10^6$	$901.7 \pm 72.2^B$	$1270.7 \pm 78.1^A$	$2046.7 \pm 65.8^{AB}$	$5.0 \pm 0.1^{AB}$
	Control	$1234.0 \pm 71.6^A$	$1105.0 \pm 111.2^{AB}$	$2341.0 \pm 53.3^A$	$5.8 \pm 0.1^A$

Different lowercase letters in the same column indicate significantly different means for isolate BIM-001, while uppercase letters in the same column indicate significantly different means for isolate BY2 ( $P \leq 0.05$ )

concentration of BY2 isolate. An inverse correlation was determined between the spore concentration density of the BY2 isolate and the total nematode density, and reproduction rate values. The lowest mean of total nematode density ( $1492.2 \pm 99.7$ /soil + root) and reproduction rate ( $3.6 \pm 0.2$ ) were found at the BY2 concentration of  $10^8$  spore/ml, while these values were found at the highest concentration of  $10^6$  spore/ml of  $2046.7 \pm 65.8$ /soil + root and  $5.0 \pm 0.1$ , respectively, and there was a statistical difference between them ( $P \leq 0.05$ ) (Table 1).

Another result obtained in the study was that the plant and root growth parameters were determined higher than the control in wheat treated with BIM-001 and BY2 isolates. Plant and root height and plant and root fresh weight reached the highest values at  $10^8$  spore/ml concentration of BIM-001 isolate compared to other concentrations. In the BY2 isolate, there was non-statistical difference between  $10^8$  and  $10^7$  spore/ml concentrations in plant and root wet weight parameters, while the values were higher at  $10^8$  spores/ml in plant and root length parameters and the statistical difference was found between  $10^8$  and  $10^7$  spore/ml concentrations in terms of these parameters ( $P \leq 0.05$ ) (Table 2). Although the root nematode density was determined higher than the control at BIM-001 and BY2  $10^6$  spore/ml concentration, there was non-significant difference in root length and wet weight parameters between the control at this concentration (Tables 1, 2).

## Discussion

In this study, the effect of concentrations of *B. bassiana* isolates on *P. thornei* varied. While the reproduction rate of *P. thornei* at  $10^8$  and  $10^7$  spore/ml concentrations in both isolates decreased than the control, at  $10^6$  spore/ml concentration was found to be similar to the control. Unfortunately, it was determined that the *P. thornei*

density in the roots increased at  $10^6$  spore/ml concentration of both isolates, and accordingly, the total nematode density and reproduction rate increased. Interestingly, a positive effect on nematode reproduction was determined at low concentration. The reason for this was that the concentration of  $10^6$  spore/ml compared to the other concentrations could not infect *P. thornei*. Since this concentration cannot reach the nematode in the soil, it is thought that the nematode density increases. Secondly, *B. bassiana* remained at a low density in the soil, substances such as the secreted enzyme or toxin may not have developed enough to kill the nematodes (Petlamul and Boukaew 2019). Therefore, the study should be repeated using higher concentrations with the same or different isolates. It is known that nematicidal activity and endophytic abilities may vary depending on the characteristics of the different fungal strains used (Karabörklü et al. 2022). In addition, detection and monitoring of *B. bassiana* in wheat root tissues was found to be necessary to improve our knowledge of the endophytic colonization of *B. bassiana*. Also, there are studies based on increasing nematode development of *B. bassiana*. Hanel (1994) hypothesized that the increased nematode numbers after the application of *B. bassiana* to the soil were due to the indirect effect of *B. bassiana*. He noted these indirect effects as higher nutrient supply for plant nematodes, changes in soil microflora, and increased nutrient availability for plants. Mwaura et al. (2017) found that ten ml of a *B. bassiana* spore suspension at a concentration of  $5 \times 10^7$  inoculated tuber and soil together with *Ditylenchus destructor* and *D. dipsaci* in the greenhouse caused a higher nematode reproduction in potatoes. Yerukala et al. (2021) reported that *B. bassiana* in Rutgers tomato cultivar increased the number of gall and egg masses formed by *M. incognita* in the roots.

**Table 2** Effects of single application of BIM-001 and BY2 isolates on *Beauveria bassiana* on wheat growth

Parameters					
Isolates	Spore Concentration	Plant length (cm)	Plant wet weight (g)	Root length (cm)	Root wet weight (g)
<i>Mean ± Standard Error*</i>					
BIM-001	$10^8$	$50.9 \pm 1.2^a$	$2.7 \pm 0.1^a$	$24.9 \pm 0.8^a$	$2.1 \pm 0.1^a$
	$10^7$	$44.8 \pm 1.0^b$	$2.0 \pm 0.1^b$	$18.6 \pm 0.8^b$	$1.6 \pm 0.1^b$
	$10^6$	$37.9 \pm 0.7^c$	$1.5 \pm 0.0^{bc}$	$16.2 \pm 0.9^b$	$0.8 \pm 0.1^c$
	Control	$35.8 \pm 1.1^c$	$1.3 \pm 0.0^c$	$17.3 \pm 0.7^b$	$1.1 \pm 0.0^c$
BY2	$10^8$	$48.1 \pm 0.8^A$	$2.4 \pm 0.1^A$	$24.0 \pm 0.7^A$	$1.7 \pm 0.0^A$
	$10^7$	$42.1 \pm 0.8^B$	$2.1 \pm 0.1^A$	$20.3 \pm 1.1^B$	$1.5 \pm 0.0^A$
	$10^6$	$36.9 \pm 0.7^C$	$1.6 \pm 0.0^B$	$17.7 \pm 0.9^B$	$1.0 \pm 0.0^B$
	Control	$35.8 \pm 1.1^C$	$1.3 \pm 0.0^B$	$17.3 \pm 0.7^B$	$1.1 \pm 0.0^B$

Different lowercase letters in the same column indicate significantly different means for isolate BIM-001, while uppercase letters in the same column indicate significantly different means for isolate BY2 ( $P \leq 0.05$ )

Additionally, few studies with different plant-parasitic nematode species support the nematicidal effect of *B. bassiana* (Ghayedi and Abdollahi 2013; Kepenekçi et al. 2017; Youssef et al. 2020; Karabörklü et al. 2022). The present study results showed that  $10^8$  and  $10^7$  spores/ml of BIM-001 and BY2 isolates negatively affected *P. thornei* soil and root density according to control. However, the most effective concentration on *P. thornei* was  $10^8$  spore/ml in both isolates. While the reproduction rate of *P. thornei* was found to be 5.8 in the control, the reproduction rates of BIM-001 and BY2 isolates in  $10^8$  spore/ml concentration were found to be 3.9 and 3.6, respectively. Although nematode reproduction rate was not suppressed at a highly, these results showed that *B. bassiana*, an entomopathogenic fungus, could control *P. thornei* infection on wheat to a certain extent. Maybe the effect on nematode control will be higher when working with higher concentrations. In addition, Göze Özdemir et al. (2022) found that *P. thornei* mortality rate was 75.5 and 64.1%, respectively, at a concentration of  $10^8$  spore/ml of *B. bassiana* BY2 and BIM-001 isolates after 72 h in vitro. The high nematicidal effect found as a result of the study carried out in vitro, unfortunately, could not be found in the experiment carried out on wheat under controlled conditions. It should not be overlooked that there was no direct contact between *P. thornei* and *B. bassiana* in the soil as in the in vitro study. Consequently, *B. bassiana*  $10^8$  and  $10^7$  spores/ml concentration applications reduced the reproduction of *P. thornei* on wheat than in the control. It was determined that these isolates showed nematicidal effect on *P. thornei*. This may be due to secondary metabolites of *B. bassiana* during the infection (Amobonye et al. 2020).

Another result of the study, when *B. bassiana* was applied to the soil contaminated with *P. thornei* at concentrations of  $10^8$  and  $10^7$  spores/ml, it was determined that the height and fresh weight of wheat plants and roots increased than the control. *B. bassiana* isolate (MARD 92) was identified to have endophytic property which enables it to be established within plant tissues and increases its field efficacy in controlling some pests (Khudhair et al. 2016). Beserra et al. (2019) reported that *B. bassiana* application in soybean positively affected vegetative development and reduced parasitism of *P. brachyurus* in roots. According to Kepenekçi et al. (2017), tomato growth and yield increased with increasing dose applications of *B. bassiana* (F-56 and F-63) isolates in tomato roots infected with *M. incognita*.

## Conclusions

In the present study, the application of native *B. bassiana* isolates BIM-001 and BY2 inoculum density was found important for biocontrol efficacy. Also, as concentration

densities above  $10^8$  spore/ml are considered more promising in control to root lesion nematodes, reassessment with different concentrations can be done. *B. bassiana* BIM-001 and BY2 isolates may be used in the control of root lesion nematode on wheat which is an important plant parasitic nematode group in Turkey and the world. Therefore, detailed research is needed on the factors responsible for the pathogenicity of *P. thornei* in the presence of the endophytic *B. bassiana* on wheat. The results of this study may guide the bionematicide recording of BIM-001 and BY2 isolates.

### Author contributions

FGGO and AUY prepared the materials, conducted the experiment, and ultimately collected the data. FGGO, AUY and OD wrote the manuscript, read and approved the final manuscript.

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### Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

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### Competing interests

The authors declare that they have no competing interests.

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## References

- Amobonye A, Bhagwat P, Pandey A, Singh S, Pillai S (2020) Biotechnological potential of *Beauveria bassiana* as a source of novel biocatalysts and metabolites. *Crit Rev Biotechnol* 40(7):1019–1034. <https://doi.org/10.1080/07388551.2020.1805403>
- Beserra AR, Almeida FA, Leite MLT, Fonseca WL, Neto FA, Pereira FF, dos Santos TS (2019) Agentes de biocontrole no manejo de *Pratylenchus brachyurus* na cultura da soja. *Revista de Ciências Agrárias* 42(3): 776–785. <https://doi.org/10.19084/rca.17201>
- Castillo P, Vovlas N (2007) *Pratylenchus* (Nematoda: Pratylenchidae): diagnosis, biology, pathogenicity and management. Brill Academic Publishers, Leiden, 555 p
- Devi G (2019) *Beauveria bassiana*: a potential biocontrol agent against plant parasitic nematodes. *Int J Curr Res* 11(1):44–47
- Ghayedi S, Abdollahi M (2013) Biocontrol potential of *Metarhizium anisopliae* (Hymenoptera: Clavicipitaceae), isolated from suppressive soils of Boyer-Ahmad region, Iran, against *J2s* of *Heterodera avenae*. *J Plant Protect Res* 53(2):165–171. <https://doi.org/10.2478/jppr-2013-0025>
- Göze Özdemir FG, Arıcı ŞE (2021) Effect of culture filtrate concentration of *Rhizoctonia solani* Kühn against *Meloidogyne incognita* and *Meloidogyne hapla* in vitro. *Int J Agric for Life Sci* 5(1):74–79
- Göze Özdemir FG, Yaşar B, Elekciöğlu H (2021) Distribution and population density of plant parasitic nematodes on cereal production areas of Isparta and Burdur Provinces of Turkey. *Turk J Entomol* 45(1):53–64. <https://doi.org/10.16970/entoted.805356>

- Göze Özdemir FG, Uzun Yiğit A, Demirözer O (2022) Assessment of nematicidal activity of *Beauveria bassiana* (Bals.-Criv.) Vuill on *Pratylenchus thornei* (Sher et Allen) (Tylenchida: Pratylenchidae). *Int J Agric Environ Food Sci* 6: 65–71. <https://doi.org/10.31015/jaefs.2022.1.10>
- Hanel L (1994) Effects of Boverol (*Beauveria bassiana*) application on soil nematodes (Nematoda) in a fallow (English abstract). *Ochrana Rostlin* 30:121–132
- Hummadi EH, Dearden A, Generalovic T, Clunie B, Harrott A, Cetin Y, Butt T (2021) Volatile organic compounds of *Metarhizium brunneum* influence the efficacy of entomopathogenic nematodes in insect control. *Biol Control* 155:104527. <https://doi.org/10.1016/j.biocontrol.2020.104527>
- Hooper DJ, Hallmann J, Subbotin SA (2005) Methods for extraction and processing. Plant parasitic nematodes in subtropical and tropical agriculture, 2nd edn. Wallingford, UK, CAB International, pp 53–84
- Karabörklü S, Aydınli V, Dura O (2022) The potential of *Beauveria bassiana* and *Metarhizium anisopliae* in controlling the root-knot nematode *Meloidogyne incognita* in tomato and cucumber. *J Asia-Pac Entomol* 25(1):101846. <https://doi.org/10.1016/j.aspen.2021.101846>
- Khoja S, Eltayef KM, Baxter I, Myrta A, Bull JC, Butt T (2021) Volatiles of the entomopathogenic fungus, *Metarhizium brunneum*, attract and kill plant parasitic nematodes. *Biol Control* 152:104472. <https://doi.org/10.1016/j.biocontrol.2020.104472>
- Khudhair MW, Alrubeai HF, Khalaf ZM (2016) Innovative method to control Dubas Bug, *Ommatissus lybicus* (Deberg) (Homoptera: Tropiduchidae) in date palm orchards using endophytic *Beauveria bassiana* isolates. *J Agric Sci Technol A* 6:394–402
- Kepenekci I, Saglam HD, Oksal E, Yanar D, Yanar Y (2017) Nematicidal activity of *Beauveria bassiana* (Bals.-Criv.) Vuill. against root-knot nematodes on tomato grown under natural conditions. *Egypt J Biol Pest Control* 27(1):117–120
- Kepenekci I, Toktay H, Oksal E, Buzboğa R, İmren M (2018) Effect of *Purpureocillium lilacinum* on root lesion nematode, *Pratylenchus thornei*. *J Agric Sci* 24(3): 323–328. <https://doi.org/10.15832/ankutbd.456647>
- Larson JE (1959) The biology of *Pratylenchus thornei* Sher and Allen, 1953. University of California, Berkeley
- Mallaiah B, Muthamilan M, Prabhu S, Ananthan R (2014) Studies on interaction of nematode, *Pratylenchus delattrei* and fungal pathogen, *Fusarium incarnatum* associated with crossandra wilt in Tamil Nadu, India. *Curr Biotica* 8(2):157–164
- Mwaura P, Niere B, Vidal S (2017) Application of an entomopathogenic fungus (*Beauveria bassiana*) increases potato nematodes reproduction and potato tubers damage caused by *Ditylenchus destructor* and *D. dipsaci*. *Biol Control* 115:23–29. <https://doi.org/10.1016/j.biocontrol.2017.08.012>
- Owen KJ, Clewett TG, Thompson JP (2010) Pre-cropping with canola decreased *Pratylenchus thornei* populations, arbuscular mycorrhizal fungi, and yield of wheat. *Crop Pasture Sci* 61(5):399–410
- Ortiz-Urquiza A, Riveiro-Miranda L, Santiago-Alvarez C, Quesada-Moraga E (2010) Insect-toxic secreted proteins and virulence of the entomopathogenic fungus *Beauveria bassiana*. *J Invertebr Pathol* 105(3):270–278. <https://doi.org/10.1016/j.jip.2010.07.003>
- Petlamul W, Boukaew S (2019) Optimisation and stabilisation of cellulase and xylanase production by *Beauveria bassiana*. *Environ Asia* 12(1):11–19. <https://doi.org/10.14456/ea.2019.2>
- Sarı HM (2020) Determination of pathogenic effects of some entomopathogenic fungus on different biological stages of Leptinotarsa decemlineata Say. Isparta University of Applied Sciences The Institute of Graduate Education Department of Plant Protection MSc Thesis 48 pp
- Söğüt MA, Yılmaz T, Göze FG, Devran Z, Elekçioğlu İH (2011) Goller Bölgesi'nde Buğday Yetiştiriciliğinde Hareketli Endoparazit Nematodlar. Türkiye IV. Bitki Koruma Kongresi Bildirileri, 28–30 Haziran 2011, Kahramanmaraş, 475, 496 (in Turkish)
- Tan JA, Jones MG, Fosu-Nyarko J (2013) Gene silencing in root lesion nematodes (*Pratylenchus* spp.) significantly reduces reproduction in a plant host. *Exp Parasitol* 133(2):166–178. <https://doi.org/10.1016/j.exppara.2012.11.011>
- Thompson JP, Sheedy JG, Robinson NA, Clewett TG (2021) Tolerance of wheat (*Triticum aestivum*) genotypes to root-lesion nematode (*Pratylenchus thornei*) in the subtropical grain region of eastern Australia. *Euphytica* 217(3):1–30. <https://doi.org/10.1007/s10681-020-02761-0>
- Thompson JP, Clewett TG, O'Reilly MM (2015) Temperature response of root-lesion nematode (*Pratylenchus thornei*) reproduction on wheat cultivars has implications for resistance screening and wheat production. *Ann Appl Biol* 167(1):1–10. <https://doi.org/10.1111/aab.12187>
- Uzun A (2020) Thysanoptera species of the cereal production areas in lakes region, distribution ratios and population fluctuations of important species and molecular characterization of some species of phlaeothripidae. Isparta University of Applied Sciences The Institute of Graduate Education Department of Plant Protection Ph.D. Thesis 266 pp
- Yerukala S, Bernard EC, Gwinn KD, Butler DM, Grewal PS, Ownley BH (2021) Endophytic *Beauveria bassiana* increases galling of 'Rutgers' tomato roots with *Meloidogyne incognita*. *J Nematol* 53:1–16. <https://doi.org/10.21307/jofnem-2021-072>
- Youssef MM, El-Nagdi WM, Lotfy DE (2020) Evaluation of the fungal activity of *Beauveria bassiana*, *Metarhizium anisopliae* and *Paecilomyces lilacinus* as biocontrol agents against root-knot nematode, *Meloidogyne incognita* on cowpea. *Bull Natl Res Centre* 44(1):1–11. <https://doi.org/10.1186/s42269-020-00367-z>
- Zuckerman BM, Mai WF, Harrison MB (1985) Plant Nematology Laboratory Manual. The University of Massachusetts Agricultural Experiment Station Amherst, Massachusetts

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