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Functional response of the predatory species *Orius albidipennis* Reuter (Hemiptera: Anthocoridae) to two life stages of *Bemisia tabaci* (Genn.) (Hemiptera: Aleyrodidae)

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Abstract

The whitefly species, *Bemisia tabaci* (Genn.) (Hemiptera: Aleyrodidae) is an important pest of many vegetable crops in the world. To evaluate the potential of the predatory species, *Orius albidipennis* Reuter (Hemiptera: Anthocoridae) in pest control, functional responses of the predator to different densities of eggs and third instar nymphs of the whitefly were determined under the laboratory conditions. Densities of (5, 8, 10, 15, 20, 25, 30, and 35) eggs and/or third instar nymphs per cucumber leaf disc were exposed to 1-day-old adults of the predator for 24-h period. The logistic regression analysis and Roger's model were used for determination of type and parameters of the predator's functional responses, respectively. The results showed that *O. albidipennis* exhibited type II and III functional responses, when fed on the whitefly eggs and third instar nymphs, respectively. The handling times (T_h) of *O. albidipennis* were calculated to be 0.35 h and 1.01 h, when it was fed on eggs and third instar nymphs of *B. tabaci*. The attack rate value (a) of the bug were calculated as 0.0723 h^{-1} on eggs, and the constant (b) was 0.0069, when it preyed on the third instar nymphs of the whitefly. The maximum attack rate (T/T_h) was calculated as 68.39 eggs and 23.20 third instar nymphs. It could be concluded that *O. albidipennis* was efficient in controlling *B. tabaci*. However, further studies in the greenhouse and field conditions are needed.

Keywords: Anthocorid bug, *Orius albidipennis*, *Bemisia tabaci*, Control potential

Background

The tobacco whitefly *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae) is a key pest of numerous host plant species found in tropical and warm temperate regions of the world (Stansly and Natwick 2010; Stansly et al. 2010). The host plants may directly experience injury from sap removal, honeydew excretion that creates favorable conditions for the rapid growth of sooty mold, which interferes with photosynthesis and also causes reduction in the product quality and acceptability, as well as physiological disorder (Stansly and Natwick 2010; Stansly et al. 2010). The whitefly also causes indirect

damage by virus transmission. For control of the pest, chemical insecticides have usually been applied by farmers, but the problems associated with chemical pesticides, including rapid development of insecticide resistance, side effect on non-target organisms, secondary pest outbreaks, environmental contaminations, and adverse effects on human health have led to the development of integrated pest management (IPM) (Pedigo 2002). Biological control has been a valuable component in IPM programs around the world for many years (Orr 2009).

Predatory bugs, belonging to the genus *Orius* (Hemiptera: Anthocoridae), are generalist predators of various soft bodied arthropods, including whiteflies (Reitz et al. 2006). *Orius albidipennis* Reuter is a common predator in several regions of Iran, and its potential as a biocontrol agent has been reported, especially under

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greenhouse conditions (Salehi et al. 2016). It is crucial to evaluate efficiency of natural enemies before using them in a biological control program (Fathipour et al. 2006). One of the criteria for evaluating a predator efficacy is its feeding response to changes in prey species, density, or its functional response (Holling 1965). Functional responses of *Orius* bugs to some pests were determined by previous researchers. For instance, functional responses of *O. insidiosus* Say to *Panonychus ulmi* Koch (McCaffrey and Horsburgh 1986), *Frankliniella occidentalis* Pergande (Coll and Ridgway 1995), and *Aphis glycines* Matsumura (Rutledge and O'Neil 2005) were studied. But, no effort has been done to determine functional response of *O. albidipennis* to *B. tabaci*.

Therefore, the present study aimed to estimate the functional response of the adult bugs to eggs and third instar nymphs of *B. tabaci* under the laboratory conditions.

Materials and methods

Host plants

Cucumber (*Cucumis sativus* cv. Servis plus) seeds were grown in plastic pots (8-cm diameter and 13-cm height) filled with a soil-cocopeat mix (1:1, v:v) and irrigated every 3 days. The pots were maintained in insect cages (120 × 100 × 60 cm) inside a controlled environment room (25 ± 5 °C, 16 L:8D). After 5–6 weeks from cultivation, the host plants were used for establishment of the whitefly colony and for trials.

Insect rearing

Adults and nymphs of *B. tabaci* were collected from cucumber fields in Mollasani region, Ahwaz, Khuzestan province, southwest of Iran. The samples were introduced into rearing cages (120 × 100 × 60 cm) containing the cucumber pots. The pots were kept inside a controlled environment room (25 ± 1 °C, 65 ± 5% RH and a photoperiod of 16 L:8D). After the establishment of the colony, eggs and third instar nymphs were used in the experiments. To obtain the cohorts, five couples of whitefly adults (each included one male and one female) were collected from the rearing stock and confined on host plant leaves by a clips cage (2.5-cm diameter). The adults were removed after 24 h. The eggs and third instar nymphs were obtained for the experimentation after 1 and 10 days, respectively.

Adults of *O. albidipennis* were collected from corn and sunflower fields in Mollasani region, Khuzestan province, southwest of Iran. The female bugs were isolated in a Plexiglas cylinder (18 cm high, 7.5cm diameter). The top of lid cylinder was covered by a fine net for ventilation as well as the holes placed on its surface. The nymphs and adults of the predatory bug were fed on frozen *Ephestia kuehniella* Zeller (Lepidoptera:

Pyralidae) eggs and pollens of corn and sunflower. The bean pods, *Phaseolus vulgaris* L., were used as oviposition substrates. The predator was reared at (25 ± 1 °C, 65 ± 5% RH, and 16:8 h (L:D)) in an environmental controlled room. For reduction of cannibalism, the rearing cylinders were lined by crumpled wipe papers.

Experimental design

The experimental arena was a Petri dish (10-cm diameter). A hole (2-cm diameter) was constructed in the fine gauze cover lid for ventilation. Inside each Petri dish, a cucumber leaf disc (5-cm diameter) was placed upside down on a wetted filter paper to keep the leaf discs fresh (Montserrat et al. 2000). Densities of (5, 8, 10, 15, 20, 25, 30, and 35) the preys (eggs or third instar nymphs of *B. tabaci*) per arena were used. The densities were determined according to preliminary tests. A female predator (1 to 5 days old), kept starved for 24 h before the experiment, was introduced to each arena. Following 24 h of exposure, the predatory bugs were removed and the numbers of consumed eggs and nymphs were recorded, separately. The trials were performed at (25 ± 1 °C, 65 ± 5% RH, and a photoperiod of 16:8 h (L: D)) in an environmentally controlled room. Ten replicates were conducted at each prey density.

Data analyses

Analysis of variance (ANOVA) by SAS 9.1 was used to compare consumed preys among different prey densities. The type of the functional response of predator was determined by logistic regression analysis of the proportion of consumed preys (N_a/N_0), as a function of prey offered (N_0) (Juliano 2001). Data were fitted to a polynomial function that describes the relationship between N_e/N_0 and N_0 (Eq. 1):

$$\frac{N_a}{N_0} = \frac{\exp(P_0 + P_1N_0 + P_2N_0^2 + P_3P_0^3)}{(1 + \exp P_0 + P_1N_0 + P_2N_0^2 + P_3N_0^3)} \quad (1)$$

where N_a is the number of consumed prey, N_0 is the initial number of preys, and P_0 , P_1 , P_2 , and P_3 are the intercept of the linear, quadratic, and cubic coefficients, respectively. These parameters were calculated, using the method of maximum likelihood (PROC CATMOD, SAS Institute, 2001). When $P_1 > 0$ and $P_2 < 0$, the proportion of consumed preys is positively density-dependent—the data describes a type III of functional response, but if $P_1 < 0$ and $P_2 > 0$, the proportion of consuming preys decreases gradually as the initial number of prey offered increases—the data indicates a type II of functional response (Juliano 2001). In the second step of the analysis, nonlinear least squares regression (PROC NLIN; SAS Institute Inc. 2001) was used to fit Roger's random attack model, which describes a type

II or III functional response (Eq 2 and Eq 3, respectively), for estimation of the functional response parameters, respectively. Because prey density was reduced during the trial, this model, which does not assume a constant prey density, is a suitable choice for the analysis (Juliano 2001).

$$N_a = N_0 \{1 - \exp[-aTP_t / (1 + aT_h N_t)]\} \quad (2)$$

where N_a is the number of consumed prey, N_0 is the initial number of prey, a is the instantaneous attack rate (searching efficiency), T is the total amount time available for searching (in this experiment $T = 24$ h), P_t the number of predators, and T_h is the handling time.

$$N_a = N_0 \frac{1 - \exp[(d + bN_0)(ThN_a - T)]}{1 + cN_0} \quad (3)$$

where b , c , and d are constants from the function that relates the attack coefficient (a) and N_0 in the type III functional response. The attack coefficient (a) is calculated by Eq 4 (Hassell 1978).

$$a = \frac{(d + bN_0)}{1 + cN_0} \quad (4)$$

The curves of the number of consumed prey by *O. albidipennis* females to different densities of the eggs and third instar nymphs of *B. tabaci* were depicted by Excel software.

Results and discussion

The results showed that at densities of 5–25 eggs or nymphs per arena, the consumed prey significantly increased with increasing prey density. But at densities more than 25 eggs or nymphs per arena, the number of consumed prey was not significantly different (Table 1).

The obtained data from the logistic regression analysis of the functional response of the females of *O. albidipennis* implicated that the predatory bug showed the type II and III functional responses, when fed on the whitefly eggs and third instar nymphs, respectively

(Table 1). Functional response curves of female adults of *O. albidipennis* to the various densities of eggs and third instar nymphs of *B. tabaci* are presented in (Figs. 1 and 2), respectively. The number of consumed eggs or nymphs by the bug increased with increasing prey density (Figs. 1 and 2).

In many cases, *Orius* spp. exhibited type II functional response (Holling, 1965); however, type III functional response shows the ability of predators to increase their rate of attack with increasing in prey density (Hassell 1978), which means that the predator learned how to overcome some hurdles with catching the prey (Schenk and Bacher 2002). Type II functional response was previously reported for *O. albidipennis*, when preyed on *Megalurothrips sjostedti* Tymbom larvae (Gitonga et al. 2002), *T. tabaci* (Thysanoptera: Thripidae) (Madadi et al. 2007), and *Tetranychus turkestanii* Ugarov & Nikolski (Acari: Tetranychidae) (Hassanzadeh et al. 2015). However, the predatory bug showed type III functional response to *Tuta absoluta* Meyreck (Lepidoptera: Gelechiidae) (Salehi et al. 2016). In contrast to the present results, type II functional responses of 4 heteropteran predators, *Dicyphus tamaninii* Wagner and *Macrolophus caliginosus* Wagner (Het., Miridae), *Orius majusculus* Reuter and *Orius laevigatus* Fieber (Het., Anthocoridae), to *Trialeurodes vaporariorum* Westwood (Homoptera: Aleyrodidae) nymphs were determined (Montserrat et al. 2000). Similar to the obtained data, *M. caliginosus* bug exhibited type III functional response to nymphs of *T. vaporariorum* (Enkegaard et al. 2001). Obtained results indicated that types and estimated parameters of the functional response of the predatory bug were different, when it was fed on eggs or nymphs of *B. tabaci*. This variation may be related to the differences in size. Size of prey is an important factor that determines the type and parameters of a predator functional response (Hassell et al. 1977 and Streams 1994). These findings are in line with the results of Gitonga et al. (2002) who demonstrated that the *O. albidipennis* showed different types and parameters of functional

Table 1 Maximum-likelihood estimates from logistic regressions of the proportion of *Bemisia tabaci* eggs and third instar nymphs eaten by *Orius albidipennis* on initial prey density

Developmental stage	Type	Coefficient	Estimate	SE	Chi-Squared value	P value
Third nymphal instar	III	Constant	-0.1938	0.6407	0.09	0.07623
		Linear	0.1083	0.1150	0.89	0.03464
		Quadratic	-0.00184	0.00592	0.10	0.07559
		Cubic	-0.00003	0.92×10^{-4}	0.08	0.07711
Egg	II	Constant	0.5908	0.6447	0.84	0.03595
		Linear	-0.0719	0.1157	0.39	0.05344
		Quadratic	0.00837	0.00597	1.96	0.01613
		Cubic	-0.000119	0.93×10^{-4}	4.05	0.0443

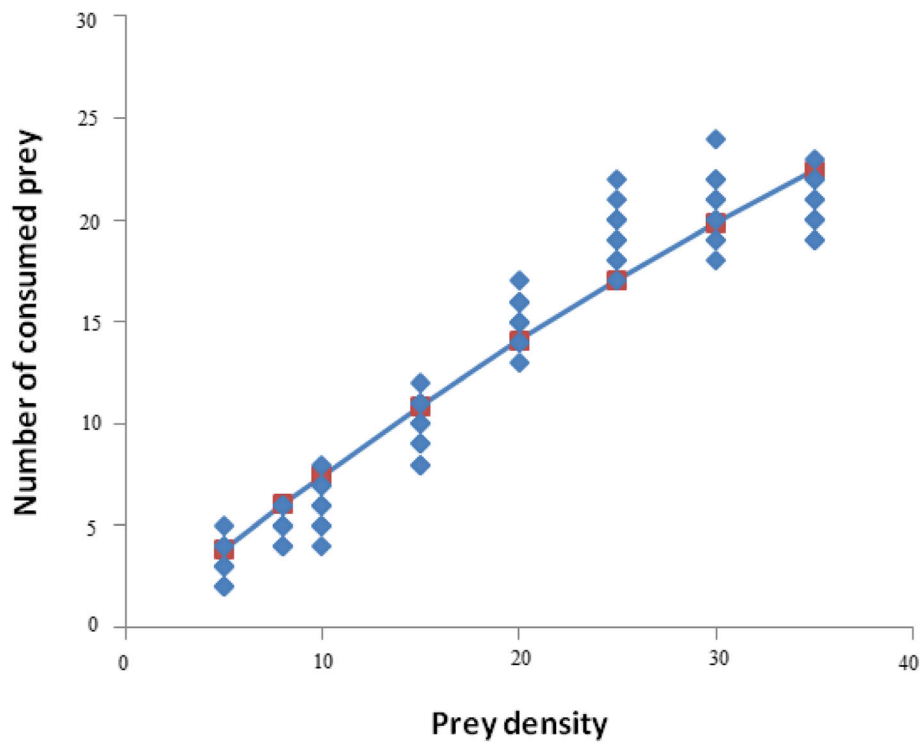


Fig. 1 Mean number of *Bemisia tabaci* eggs preyed upon by females of *Orius albidipennis* in 24 h as a function of egg density

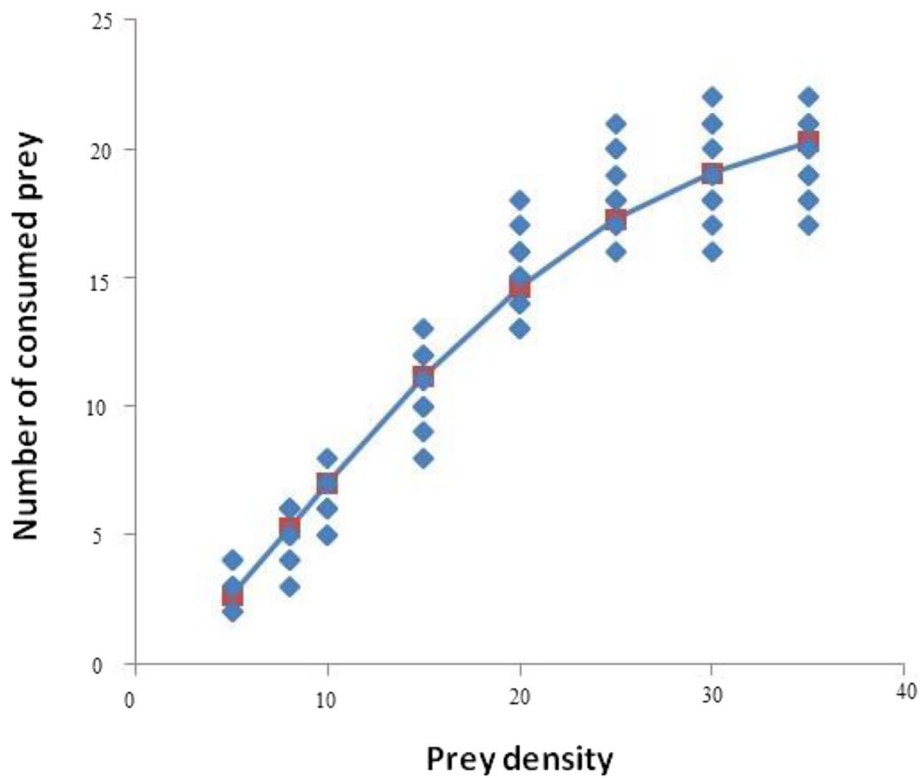


Fig. 2 Mean number of *Bemisia tabaci* third instar nymphs preyed upon by females of *Orius albidipennis* in 24 h as a function of nymph density

Table 2 Estimated (mean \pm SE) searching efficiency (a) and handling time (T_h) of *O. albidipennis* female on eggs and third nymphal stages of *B. tabaci*

Developmental stage	Parameters	Estimate	Asymptotic SE	Asymptotic 95% CI		T/T_h	R^2
				Lower	Upper		
Third nymphal instar	Constant (b)	0.0069	0.000612	0.00598	0.00842	23.20	0.9551
	T_h	1.034	0.0314	0.9519	1.0772		
Egg	a	0.0723	0.00693	0.0506	0.0782	68.39	0.997
	T_h	0.3509	0.0975	0.1667	0.5548		

response, according to its prey life stages. Functional response models often assume the homogenous prey size and size-independent feeding rates. However, variation in prey size due to ontogeny and competition is ubiquitous, and predation rates are often size dependent (McCoy et al. 2011).

Estimated functional response parameters, when the bug preys on eggs or third instar nymphs of the pest, are shown in Table 2. The handling times (T_h) of *O. albidipennis* were calculated to be 0.350 h and 1.034 h, when fed on eggs and third instar nymphs of *B. tabaci*, respectively. The attack rate value (a) of the bug was calculated as 0.0723 h^{-1} on eggs, and the constant (b) was 0.0069, when it preyed on the third instar nymphs of the whitefly. The maximum attack rate (T/T_h) was calculated as 68.39 eggs and 23.20 third instar nymphs (Table 2). The calculated values of T_h and a in the present study were different from the values reported for *O. majusculus* and *O. laevigatus* to *T. vaporarium* (Montserrat et al. 2000). The difference may be due to different prey and predator species.

Conclusion

Determination of the functional responses of *O. albidipennis* under laboratory conditions, using simple arenas and a single prey type, may be of a limited value to predict entire predator's behavior under field or greenhouse conditions. Therefore, further studies in the greenhouse and field conditions are needed.

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Availability of data and materials

Not applicable

Authors' contributions

FY and NZS designed the experiments. AS performed the experiments. FY and NZS analyzed the data. FY wrote the manuscript. The manuscript studied and certificated by all authors. All authors read and approved the final manuscript.

Ethics approval and consent to participate

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Consent for publication

Not applicable

Competing interests

The authors declare that they have no competing interests.

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